ORIGINAL CONTRIBUTION

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An investigation of the inhibitory effects of dichloromethane and methanol extracts of Salvia macilenta, Salvia officinalis, Salvia santolinifola and Salvia mirzayanii on diabetes marker enzymes, an approach for the treatment diabetes



Abstract

Background: Diabetes mellitus is a type of metabolic disease characterized by elevated blood sugar. The main strategy for its treatment is to inhibit carbohydrate-hydrolyzing enzymes, including α-amylase and α-glucosidase. The aim of the present study was to evaluate the efficacy of *Salvia* extracts in inhibiting diabetes marker enzymes.

Materials and methods: This experimental study was performed in vitro. The studied plants included *Salvia mirzayanii*, *Salvia macilenta*, *Salvia officinalis* and *Salvia santolinifola* and inhibitory effects of their methanolic and dichloromethane extracts were investigated. After calculating the percentage of α -amylase inhibition and IC₅₀ of the extracts, Km and Vmax were also determined using prism7. Statistical analysis was performed employing with Graph Pad instat3 software.

Results: The results here in revealed that methanol extracts of *Salvia santolinifola* (with IC₅₀ = 54.72 \pm 9.6 μg / ml) and *Salvia officinalis* with (IC₅₀ = 54.87 \pm 5.7 μg / ml) and dichloromethane extract of *Salvia officinalis* with (IC₅₀ = 71.20 \pm 14.3 μg / ml) had the greatest inhibitory effect on α-amylase comparing to acarbose with (IC₅₀ = 42.94 \pm 3.8 μg / ml) as a standard. Tukey test results showed that there is a significant difference between IC₅₀ of acarbose comparing to methanol extract of *Salvia mirzayanii* and dichloromethane extracts of *Salvia mirzayanii* and *Salvia santolinifola* with *P* value 5 0.001 in α-amylase inhibition.

Conclusion: The extracts had significant inhibitory effects on α -amylase inhibition. Among the extracts of the studied species, methanol extract of *Salvia santolinifola* demonstrated the greatest inhibitory effect on α -amylase.

Keywords: Diabetes, α-Amylase, Salvia extracts, Inhibition percentage, Kineticof inhibition

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Background

Diabetes mellitus is one of the most important public health problems in the world. According to the International Diabetes Federation reports, there are 415 million adults with diabetes, which is likely to reach 612 million by 2040 [1]. Unless action is taken, it is predicted that by 2030 at least 350 million people will have type 2 diabetes.



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Some Lamiacea, such as Salvia mirzayanii, are used to treat diabetes [2]. Diabetes mellitus is an endocrine and metabolic disorder characterized by chronic hyperglycemia caused by adefect in insulin secretion or its function. Diabetic patients in many countries around the world suffer from postprandial hyperglycemia (PPHG) due to changes in their diet and lifestyle and high carbohydrate intake. This can progress to type 2 diabetes. Current strategies for the treatment and control of postprandial hyperglycemia included inhibition of α-glucosidase and α-amylase resulting in delayed digestion and absorption of carbohydrates. α -amylase and α -glucosidase inhibitors such as acarbose and voglibosecan potentially reduce the progression of the diabetes and prevented from the neuropathy, nephropathy, and retinopathy. However, these drugs have side effects such as diarrhea, abdominal pain, and gastrointestinal complications; therefore, many efforts have been made to replace gastrointestinal inhibitors [3].

Glucosidases are a group of carbohydrate digestive enzymes that break down carbohydrates into monosaccharides. A number of traditional Iranian medicinal plants, especially Salvia mirzayanii(S.mirzayanii), act as α-amylase inhibitor in the prevention and management of diabetes [4]. Inhibition of α -amylase and α -glucosidase specifically reduced postprandial hyperglycemia and thus prevented the progression of diabetes [5]. Vegetables and plants have been consumed by humanas food and for the treatment of degenerative diseases. Scientific evidence proves that many of these herbs and herbal materials have medical properties that reduce symptoms of the diseases and prevent from these diseases. Research on medicinal plants for diabetes management in recent years was the interest of specialists and it has been found that some of them have anti-hyperglycemic properties, acting by the inhibition of carbohydrate hydrolyzing enzymes. This property is due to the presence of biological active compounds such as phenols and flavonoids. Phenols also have antioxidant properties due to hydroxyl groups [6].

Salvia is the largest genus of Lamiacea with more than 900 species, spreading throughout the world, especially in the temperate and warm areas. In Iran, Salvia contains 58 species, 17 of which are endemic. Extracts of Salvia species have various biological activities including antioxidant, antibacterial, anti-tumor, anti-inflammatory and anti-diabetic. In traditional Iranian medicine, some species of Salvia have antidiabetic effects [7]. Medicinal plants have always been an important source of natural compounds with high therapeutic potential [8]. The first goal of diabetic patients treatment is to control hyperglycemia [9]. In this study, we investigated α-amylase inhibition by different species of Salvia, as an approach for the treatment of diabetes.

Materials and methods

3, 5 dinitro salicylic acid (DNSA) and α -amylase enzyme from porcine pancreas and acarbose were purchased from Sigma Aldrich USA and other chemicals were obtained from Merck Company in Germany.

Specimen collection and preparation

Salvia mirzayanii (S.mirzayanii), Salvia officinalis (S.officinalis), Salvia macilenta (S.macilenta) and Salvia santolinifola (S.santolinifola) plants were collected by Mr. Asadipourin March 2018, from around Bandar Abbas. The voucher numbers were deposited in Medicinal Plants Processing Research Center, School of Pharmacy, Shiraz University of Medical Sciences. The voucher numbers of S.mirzayanii, S.santolinifola, S.officinalis and S.macilenta are MPPRC-93-6, MPPRC-97-2, MPPRC-90-18 and MPPRC-93-1, respectively. The leaves of all plants were dried, chopped, and extracted with dichloromethane and methanol using a rotary evaporator, respectively. After that, Salvia extracts were prepared and stored at 4°C.

The inhibition of α -amylase (EC:3.2.1.1) was evaluated with minor modifications by concentrations of 0.0025, 0.1,0.2 and 0.5 mg / ml dichloromethane and methanol extracts of *S.mirzayanii*, *S.macilenta*, *S. officinalis* and *S. santolinifola* according to the method reported by Maccu and Shetty, [10]. In this method, the amount of inhibition is determined using starch (45 λ ,1%) as a substrate and 3,5 dinitro salicylic acid as a reagent. First, *Salvia* extracts (45 λ) are mixed with distilled water and starch. The reaction begins with the addition of α -amylase (10 λ) and the same amount of sodium phosphate buffer (PH=6.9) was added. The mixture was incubated at 37 °C for 15 min and 100 λ of dinitro salicylic acid and sodium potassium tartrate (as stop solution) was added.

After 15 min incubation at 85 °C, the activity of α -amylase was measured by measurement of absorbance at 540 nm (Biotek Corporation's synergy HTX multi-mode reader). To measure enzyme inhibition by the control, the above mentioned procedure was performed, only DMSO (*Dimethyl sulfoxide*) is used instead of the *Salvia* extract. In this study, acarbose is employed as a standard.

The percentage of enzyme inhibition by the samples and acarbose was determined by the following formula.

%I = 100-((OD control-OD sample) /OD control) × 100. Each experiment was repeated 3 times .

Blank contained sodium phosphate buffer instead of the enzyme. We calculated the IC_{50} of α -amylase inhibition by *Salvia* extracts. Then Km and Vmax of the enzyme were calculated.

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Kinetics of α-amylase enzyme

To determine the type of inhibition and to calculate Km and Vmax, different concentrations of substrate (starch) (0.25%, 0.5%, 1%, 2%, 4% and 6%) were prepared. The extract concentration, which caused 50% inhibition, was used. The experiment was started with a low concentration of substrate, (starch). The experiment was started (according to the mentioned procedure described previously) with low substrate concentration until Michaelis Menten curve reaches saturation or plateau (this plateau showed maximum velocity). Then, Km and Vmax were calculated using Graph pad prism software. Comparison of Km and Vmax in the absence of inhibitor and in the presence of inhibitor indicates the type of inhibition.

Statistical analysis

After calculating IC₅₀ by Excel 2019 software, Graph Pad prism 7 and Graph Pad instat 3 softwares were used for statistical analysis. Mean \pm SD values of the obtained data were calculated using Graph Pad instat3 and One-way ANOVA with Tukey posttest. If P value was $^{<}$ 0.05 then the difference is considered as significant. Graph Pad prism6 software was also used to calculate Km and Vmax values.

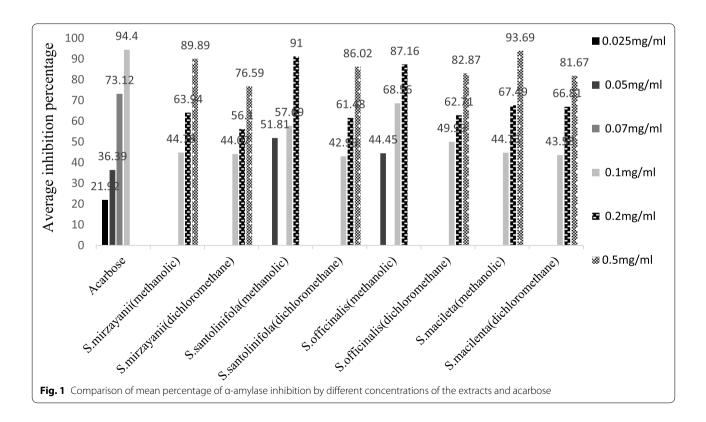
Results

The average percentage of α -amylase inhibition by *Salvia* species and acarbose was determined. Figure 1 shows a comparison of the mean percentage of α -amylase inhibition. As shown, acarbose had the highest inhibition rate (94%) as a standard at 0.1 mg / ml. In addition, at the concentration of 0.2 mg / ml, methanol extract of *S.santolinifolia* with 91% and methanol extract of *S.officinalis* with 87.1% showed the highest inhibition percentage. At 0.5 mg / ml concentration, the inhibitory effect of methanol extract of *S.macilenta* (93.69%) was higher than other species.

The results of the calculated IC₅₀ (the concentration of *Salvia* extract which inhibits α -amylase 50%) \pm SD by GraphPad instat3 software are presented in Table 1 with respect to repeated experiments.

Concentration and activity values were then entered in Graphpad prism software and Km and Vmax values were calculated. The results of Km \pm SD and Vmax \pm SD are presented in Table 2.

Comparison of Km and Vmax with and without inhibitor determines the type of inhibition. The inhibition of the enzyme by acarbose, is competitive (Vmax is constant and Km increased). The inhibition of the enzyme by *Salvia* extracts is mixed, (Vmax decreased and Km increased).



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The saturation diagram (Michaelis Menten) of the α -amylase for the one-time trial drawn by Excel 2019 software and was shown in Figs. 2 and 3.

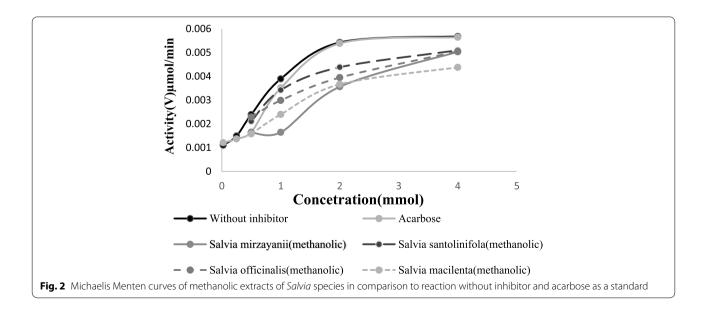
As is known, the activity without inhibitor was higher than that of the studied Salvia species. To convert absorbance to the activity, maltose standard diagram was drawn. Then, we calculate the amount of activity using the line equation of maltose standard diagram. (The standard maltose curve with four points for converting OD to activity is shown in Fig. 4). Then, concentration and activity values were entered in Graphpad prism software and values of Km and Vmax were calculated.

Table 1 $IC_{50} \pm SD$ of the extracts and acarbose (µg/ml)

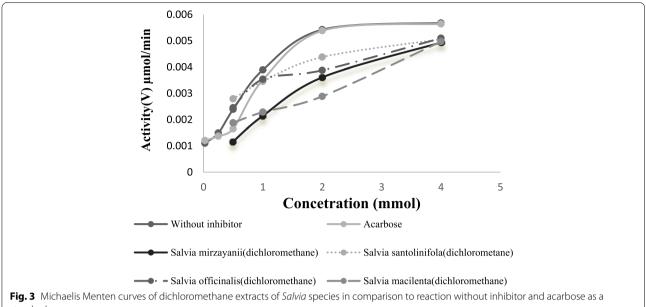
Samples	α -amylase inhibition (IC ₅₀ , μg/ml)	Samples	α-amylase inhibition (IC ₅₀ , μg/ml)
S. mirzayanii (methanolic)	114.8 ± 11.1	S. officinalis (methanolic)	54.9 ± 5.7
S. mirzayanii (dichloromethane)	150.9 ± 6.7	S. officinalis (dichloromethane)	71.2 ± 14.4
S. santolinifola (methanolic)	54.7 ± 9.6	S. macilenta (methanolic)	103.7 ± 1.1
S. santolinifola (dichloromethane)	132.9 ± 15.8	S. macilenta (dichloromethane)	100.9 ± 10.8
Acarbose	42.9 ± 3.9		

Table 2 Km \pm SD and Vmax \pm SD and the mode inhibition of the studied samples and acarbose

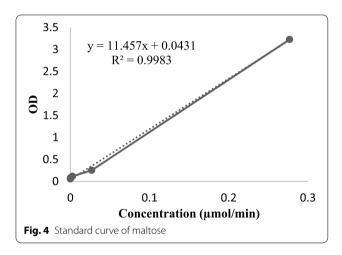
Extract	Km ± SD)mmol(Vmax±SD)μmol/min(Kind of inhibition
Acarbose (standard)	1.264±0.301↑	0.0076 ± 0.0007(constant)	Competitive
Salvia mirzayanii (methanolic)	1.969±0.142↑	$0.0075 \pm 0.0002 \downarrow$	Mixed
Salvia mirzayanii (dichloromethane)	2.398±0.736↑	$0.0074 \pm 0.0009 \downarrow$	Mixed
Salvia santolinifola (methanolic)	1.351 ± 0.208↑	$0.0075 \pm 0.0003 \downarrow$	Mixed
Salvia santolinifola (dichloromethane)	1.082 ± 0.242↑	0.0073 ± 0.0004	Mixed
Salvia officinalis (methanolic)	1.665 ± 0.219↑	$0.0075 \pm 0.0003 \downarrow$	Mixed
Salvia officinalis (dichloromethane)	1.343 ± 0.245↑	0.0075 ± 0.0004	Mixed
Salvia macilenta (methanolic)	2.339±0.315↑	0.0074 ± 0.0004	Mixed
Salvia macilenta (dichloromethane)	2.456 ± 0.541↑	$0.0073 \pm 0.0006 \downarrow$	Mixed
Without inhibition	1.01 ± 0.213	0.0076 ± 0.0006	=



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standard



Tukey post hoc test, compares the IC₅₀ of the studied species and acarbose. Based on these results, there are significant differences between the IC₅₀ of acarbose and methanol, and dichloromethane extracts of S.mirzayanii and dichloromethane extract of *S.santolinifola* (P value < 0.001) in α -amylase inhibition. Also, there was not a significant difference between methanol, dichloromethane extracts of S.officinalis and acarbose (P value> 0.05). Also, there is a significant difference between the of IC50 of acarbose, methanol and dichloromethane extracts of S.macilenta (P value < 0.01). Between different samples, methanolic extract of S.officinalis (IC₅₀ = $54.9 \pm 5.7 \,\mu\text{g/ml}$) and methanolic extract of S.santolinifola (IC₅₀ = $54.7 \pm 9.6 \,\mu\text{g}$ /

ml) possessed the highest α -amylase inhibition, respectively.

Discussion

Type 1 diabetes is insulin dependent and type 2 diabetes is non-insulin dependent. About 90% of people suffer from type 2 diabetes. Although, with insulin injection, type 1 diabetes is treatable, finding drugs to treat and manage type 2 diabetes is an important area of research. Pancreatic α-amylase and intestinal α-glucosidase are key enzymes that convert carbohydrates to monosaccharides. Therefore, they are responsible for elevation blood sugar; and inhibiting these enzymes is an appropriate way to treat diabetes [11]. The α -amylase enzyme has three domains (A, B, C) and has 496 amino acids. The catalytic site of α-amylase (PA) contains Asp197-Glu223-Asp300 amino acids [12]. Salvia has been so popular that it has been the subject of numerous chemical studies. These plants are rich in polyphenols and contain more than 160 polyphenols, some of which are unique to the genus Salvia. Several phenolic acetophenone glycosides have only been identified in S. officinalis [13]. There is a strong correlation between plant phenol content and inhibitory activities of α -amylase and α -glucosidase. Polyphenols are considered as potent antioxidants due to their redox properties in their hydroxyl groups [14]. One of the important functions of polyphenols is to inhibit digestive enzymes, especially carbohydrate hydrolyzing enzymes such as α -amylase and α -glucosidase. Inhibitors of enzymes capable of digestion carbohydrates decrease glucose absorption. Polyphenolic α -amylase and Javid et al. Clinical Phytoscience (2022) 8:7 Page 6 of 8

 α -glucosidase inhibitors derived from natural sources have been reported to be beneficial in reducing postprandial hyperglycemia [15].

The results of this study showed that dichloromethane and methanol extracts of S.mirzayanii inhibit α-amylase. In addition, according to Table 1, their mean IC_{50} ($IC_{50} = 150.9 \pm 6.7 \,\mu\text{g} / \text{ml}$, $IC_{50} = 114.8 \pm 11.1 \,\mu\text{g} /$ ml) respectively,is higher than the standard IC50 acarbose (IC₅₀ = $42.9 \pm 3.9 \,\mu\text{g}$ / ml). In other words, their levels of α-amylase inhibition were lower than acarbose. These results are consistent with the findings of a study by Dr. Moein et al. (2012) which showed that phenolic compounds in *S.mirzayanii* may be responsible for free radical scavengingand the prevention of disease such as cancer and cardiovascular disease, diabetes and aging [16]. It is also in line with Dr. Moein's 2018 study, which showed that aqueous-methanol fractions of *S.mirzayanii* inhibited the α-amylase, but further studies were needed to determine the type of inhibition [17]. Study by Mohammad Reza Zarshenas in 2014 is consistent with the above mentioned results on diabetic model of rats, which showed that injection alcoholic extractof S.mirzayanii decrease blood glucose levels [18]. The results of this study also showed that dichloromethane (IC₅₀ = $71.2 \pm 14.4 \,\mu\text{g}$ / ml, Table 1) and methanol (IC $_{50} = 54.9 \pm 5.7 \, \mu \text{g/ml}\text{,}$ Table 1) extracts of S.officinalis lower than acarbose $(IC_{50} = 42.9 \pm 5.7 \,\mu g/ml)$. These results are consistent with the findings of Ahmad Ghorbani et al. (2016), who identified the phytochemical compounds of S. officinalis flowers, leaves and stems. These compounds included alkaloids, carbohydrates, fatty acids, glycoside derivatives such as flavonoid glycosides, phenolic compounds and terpenoids that had anti-mutagenic, anti-inflammatory effects and anti-hypoglycemic activity and increased memory [19]. More than 120 compounds have been characterized in the essential oil of aerial parts of S. officinalis. The major components of this oil include borneol, camphor, caryophyllene, cineole, elemene, humulene, ledene, pinene, and thujone. Oxidative stress plays a significant role in the initiation and progression of several diseases, such as diabetes. The most effective antioxidant components of S. officinalis are carnosol, rosmarinic acid, carnosic acid, caffeic acid, rosmanol, rosmadial, genkwanin, and cirsimaritin [19]. It is also in line with studies by Rahman Mahdizadeh et al. (2018) who stated S.officinalis reduces blood sugar levels in mice and inhibits gluconeogenesis and glycogenolysis in the liver [20]. S. officinalis decreases blood glucose like acarbose, and also inhibits the activity of the intestinal maltase and sucrase and therefore possesses antidiabetic activity [20]. These results are in line with the results of Eidey et al. (2009) who reported that methanolic extract of *S.officinalis* significantly reduced blood glucose in hyperglycemic rats. In other study, ethanol extract of *S.officinalis* leaves decreased blood glucose in rat diabetic model [21].

The results of this study showed that dichloromethane (IC $_{50}=100.9\pm10.8\,\mu g$ / ml, Table 1) and methanol (IC₅₀ = $103.7 \pm 1.1 \,\mu g$ / ml, Table 1) extracts of S.macilenta inhibited α-amylase, lower than acarbose $(IC_{50} = 42.9 \pm 3.9 \,\mu\text{g} / \text{ml}, \text{ Table 1})$. Vasil Georgiev et al. (2017) reported that S.macilenta is rich in phenolic acid and polyphenols, flavonoids and terpenoids. These compounds have hypoglycemic properties [22]. It is also Mujtaba Asadullah's reported that (2017) extracts of S.macilenta and S.officinalis are used in traditional Iranian medicine for treating many disorders such as diabetes [23]. The results of this study showed that dichloromethane (IC₅₀ = $132.9 \pm 15.88 \,\mu g$ / ml, Table 1) and methanol (IC₅₀ = $54.7 \pm 9.6 \,\mu g$ / ml, Table 1) extracts of S.santolinifola inhibited α-amylase, lower than acarbose (IC₅₀ = $42.9 \pm 3.9 \,\mu\text{g}$ / ml, Table 1).

Also, S.Asadietal. (2011) reported that DNA damage by free radicals is inhibited by *Salvia* species [24]. Dichloromethane solvent extracts diterpenes, and methanol solvent extracts polyphenols. Polyphenols are generally presented in plants, and these polyphenolic compounds have been reported to be α -amylase inhibitors [4].

Based on Table 2, acarbose showed competitive inhibition with $(Km=1.264\pm0.301 \text{ and } Vmax=0.0076\pm0)$ compared with no inhibitors with $(Km=1.01\pm0.213 \text{ and } Vmax=0.0076\pm0.0006)$.

All the studied extracts showed mixed inhibition and Km and Vmax of these samples are shown in Table 2.

Uncompetitive and non-competitive inhibitors have the ability to inhibit the enzyme more than acarbose as a competitive inhibitor and, unlike acarbose, their inhibitory potency is not affected by substrate concentration [25, 26]. In 2017, Dr. Homaei and Soodeh Bahramian used the Michaelis Menten equation to determine Km, Vmax of plant extractsand type of α -amylase inhibition [27].

Conclusion

The extracts had significant inhibitory effects on α -amylase. Among the extracts of the studied species, methanolic extracts of *S.santolinifola* and *S.officinalis* had moreinhibitory effects on α -amylase. The type of inhibition of *Salvia* species, is mixed with decreasing Vmax and increasing Km, and the studied samples could inhibit α -amylase independent of the substrate concentration. However, the type of acarbose inhibition is competitive, which is affected by the substrate concentration.

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Abbreviations

 IC_{50} : the concentration of the extract inhibits enzyme 50%; kg: kilogram; ml: millimeter; ANOVA: Analysis of Variance; C: Centigrade degree; g: gram; Km: Michaelis Menten constant; Vmax: MaximumVelocity; S: Salvia.

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Authors' contributions

SM and MM designed the study. SM helped in writing research paper and revised the final manuscript. SM statistically analyzed the obtained data. HJ carried out all parts of the experiments and writing the research paper. MM was involved in selection of plants and their extraction. This research was supervised by SM. The authors read and approved the final manuscript.

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Availability of data and materials

All the data is contained in the manuscript.

Declarations

Ethics approval and consent to participate

The study protocol was approved by institutional ethical committee (No:IR. HUMS.REC.2018.270).

Consent for publication

Not applicable.

Competing interests

The authors declare that there are no conflicts of interest for publication this manuscript.

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References

- Proença C, Freitas M, Ribeiro D, Oliveira EF, Sousa JL, Tomé SM, et al. α-Glucosidase inhibition by flavonoids: an in vitro and in silico structure– activity relationship study. J Enzyme Inhib Med Chem. 2017;32(1):1216– 28. https://doi.org/10.1080/14756366.2017.1368503.
- Rouzbehan S, Moein S, Homaei A, Moein MR. Kinetics of a-glucosidase inhibition by different fractions of three species of Labiatae extracts: a new diabetes treatment model. Pharm Biol. 2017;55(1):1483–8. https://doi.org/10.1080/13880209.2017.1306569.
- Adisakwattana S, Ruengsamran T, Kampa P, Sompong W. In vitro inhibitory effects of plant-based foods and their combinations on intestinal α-glucosidase and pancreatic α-amylase. BMC complementary Altern Med. 2012;12(1):110. https://doi.org/10.1186/1472-6882-12-110.
- Moein S, Pimoradloo E, Moein M, Vessal M. Evaluation of antioxidant potentials and α-amylase inhibition of different fractions of labiatae plants extracts: as a model of antidiabetic compounds properties. Biomed Res Int. 2017;2017 https://core.ac.uk/download/pdf/204491186. pdf.
- Sheliya MA, Begum R, Pillai KK, Aeri V, Mir SR, Ali A, et al. In vitro α-glucosidase and α-amylase inhibition by aqueous, hydroalcoholic,

- and alcoholic extract of Euphorbia hirta L. Drug Dev Ther. 2016;7(1):26. https://doi.org/10.4103/2394-6555.180156.
- Ademiluyi AO, Oboh G. Aqueous extracts of Roselle (Hibiscus sabdariffa Linn.) varieties inhibit α-amylase and α-glucosidase activities in vitro. J Med Food. 2013;16(1):88–93. https://doi.org/10.1089/jmf.2012.0004.
- Asghari B, Salehi P, Sonboli A, Ebrahimi SN. Flavonoids from Salvia chloroleuca with α-Amylsae and α-glucosidase inhibitory effect. Iran J Pharm Res, IJPR. 2015;14(2):609 https://www.ncbi.nlm.nih.gov/pmc/articles/ PMC4403079/
- Malki S, Abidi L, Hioun S, Yahia A. Variability of phenolic contents in ethanolic extracts of Teucrium polium L. populations and effect on antioxidant and antimicrobial activities. Biotech Res. 2015;5(4):21–7 https:// www.semanticscholar.org.
- He L, Meng S, Germain-Lee EL, Radovick S, Wondisford FE. Potential biomarker of metformin action. J Endocrinol. 2014;221(3):363. https://doi. org/10.1530/JOE-14-0084.
- Mccue P, Kwon YI, Shetty K. Anti-amylase, anti-glucosidase and antiangiotensin i-converting enzyme potential of selected foods. J Food Biochem. 2005;29(3):278–94. https://doi.org/10.1111/j.1745-4514.2005. 00020.x.
- 11. Eskandani M, Babak Bahadori M, Zengin G, Dinparast L, Bahadori S. Novel natural agents from Lamiaceae family: an evaluation on toxicity and enzyme inhibitory potential linked to diabetes mellitus. Curr Bioact Compd. 2016;12(1):34–8 https://www.ingentaconnect.com/.
- Martinez-Gonzalez Al, Díaz-Sánchez ÁG, Rosa LA, Vargas-Requena CL, Bustos-Jaimes I. Polyphenolic compounds and digestive enzymes: in vitro non-covalent interactions. Molecules. 2017;22(4):669. https://doi.org/10. 3390/molecules22040669.
- 13. Lu Y, Foo LY. Polyphenolics of Salvia—a review. Phytochemistry. 2002;59(2):117–40. https://doi.org/10.1016/S0031-9422(01)00415-0.
- Saliu J, Ademiluyi A, Akinyemi A, Oboh G. In vitro antidiabetes and antihypertension properties of phenolic extracts from bitter leaf (Vernonia amygdalina Del.). J food. Biochem. 2012;36(5):569–76. https://doi.org/10. 1111/j.1745-4514.2011.00576.x.
- Acevedoa, F,Rubilara M, Palmaa B, Shenea C. Mode of Inhibition of α-Glucosidase and α-Amylase by Polyphenol-Enriched Extracts of Maqui (Aristotelia chilensis). 2011; https://www.semanticscholar.org/paper/.
- Moein S, Moein M, Khoshnoud MJ, Kalanteri T. In vitro antioxidant properties evaluation of 10 Iranian medicinal plants by different methods. Iran Red Crescent Med J. 2012;14(12):771. https://doi.org/10.5812/ircmj.1408.
- Moein S, Jahanshai S, Rahimzadeh M, Moein M. Kinetic of α-amylase and comparison its inhibition by ethanol and aqueous extracts of Otostegia persica, Salvia mirzayanii and Zataria multiflora. Iran J Sci Technol. 2018;42(2):339–45. https://doi.org/10.1007/s40995-016-0043-6.
- Zarshenas MM, Krenn L. Phytochemical and pharmacological aspects of Salvia mirzayanii Rech. f. & Esfand. J Evid Based Complementary Altern Med. 2015;20(1):65–72. https://doi.org/10.1177/2156587214553938.
- Ghorbani A, Esmaeilizadeh M. Pharmacological properties of Salvia officinalis and its components. J Tradit Complement Med. 2017;7(4):433–40. https://doi.org/10.1016/j.jtcme.2016.12.014.
- Mahdizadeh R, Moein S, Soltani N, Malekzadeh K, Mahmoodreza M. Study the molecular mechanism of salvia species in prevetion of diabete. IJPSR. 2018;9(11):4512–21 http://eprints.hums.ac.ir/6219/2/JPSR.pdf.
- Eidi A, Eidi M. Antidiabetic effects of sage (Salvia officinalis L.) leaves in normal and streptozotocin-induced diabetic rats. Diabetes Metab Syndr. 2009;3(1):40–4. https://doi.org/10.1016/j.dsx.2008.10.007.
- Georgiev V, Pavlov A. Salvia Biotechnology: Springer; 2017. https://doi. org/10.1007/978-3-319-73900-7.
- Asadollahi M, Firuzi O, Jamebozorgi FH, Alizadeh M, Jassbi AR. Ethnopharmacological studies, chemical composition, antibacterial and cytotoxic activities of essential oils of eleven Salvia in Iran. J Herb Med. 2018;100250. https://doi.org/10.1016/j.hermed.2018.11.006.
- Asadi S, Khodagholi F, Esmaeili MA, Tusi SK, Ansari N, Shaerzadeh F, et al. Chemical composition analysis, antioxidant, antiglycating activities and neuroprotective effects of S. choloroleuca, S. mirzayanii and S. santolinifolia from Iran. Am J Chin Med. 2011;39(03):615–38. https://doi.org/10. 1142/S0192415X1100907X.
- Ghadyale V, Takalikar S, Haldavnekar V, Arvindekar A. Effective control of postprandial glucose level through inhibition of intestinal α- glucosidase by Cymbopogon martinii (Roxb.). J Evid Based Complementary Altern Med. 2012;2012. https://doi.org/10.1155/2012/372909.

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- 26. Zhang H, Wang G, Dong J. Inhibitory properties of aqueous ethanol extracts of propolis on α -glucosidase. J Evid Based Complementary Altern Med. 2015;2015. https://doi.org/10.1155/2015/587383.
- Nasab SB, Homaei A, Karami L. Kinetic of α-amylase inhibition by Gracilaria corticata and Sargassum angustifolium extracts and zinc oxide nanoparticles. Biocatal Agric Biotechnol. 2020;23:101478. https://doi.org/ 10.1016/j.bcab.2019.101478.

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